



Sanger Samples Preparations

Use only PCR tubes, strips or plates.

Plasmid samples:

- Take 150 ng-300 ng of plasmid in a maximum volume of 7 μL .

Example: When the plasmid concentration is 70ng/ μL , calculate the required volume using this formula:

$$\frac{150[\text{ng}]}{C[\frac{\text{ng}}{\mu\text{L}}]} = V[\mu\text{L}] \quad \text{e.g.,} \quad \frac{150 \text{ ng}}{70 \frac{\text{ng}}{\mu\text{L}}} = 2.14 \mu\text{L}.$$

Add 2.2 μL of the purified plasmid to a PCR tube.

- If the volume required exceeds 7 μL , add 7 μL .
(Note: This situation might lead to low-quality sequencing results).
- Add 0.5 μL primer (10 μM stock)
- Complete the volume to 7.5 μL with PCR-grade water.
- Clearly write the reaction name on the tube cap sides.

PCR products:

- Calculate the required amount of PCR product ng using this formula:

$$\frac{0.015 \cdot \text{length of PCR product [bp]}}{\text{sample concentration} [\frac{\text{ng}}{\mu\text{L}}]} = 0.135$$

Example: For a PCR product of size **450** bp at a concentration of **50** ng/ μL , the

calculation is: $\frac{0.015 \cdot 450 [\text{bp}]}{50 [\frac{\text{ng}}{\mu\text{L}}]} = 0.135 \mu\text{L}$.

If the required volume is too small to pipette, dilute the samples 1:10 and take 1.35 μL .

* If you use Exozap or any other enzymatic cleanup kit, use 1 μL of the PCR product.

- Add 0.5 μL of primer (10 μM stock)
- Complete the volume to 7.5 μL with PCR-grade water.
- Clearly write the reaction name on the tube cap and sides.



Calculate primers Tm:

* If multiple samples have Tm values that fall within the same temperature range in the table, group them in strips or plates instead of single tubes.

40 - 44.9 °C

45 - 49.9 °C

50 - 54.9 °C

55 - 60 °C

Above 60 °C

For each primer you use, calculate the melting temperature (Tm) using the following calculator in the link:

- Go to <http://insilico.ehu.es/tm.php>
 - Add your primer sequence into the top line.
 - Select the second option: **Base-Stacking Tm**
 - Adjust the following parameters as shown in the screenshot below.
 - Click on **Compute Tm** button at the top of the page.
 - Your Tm value will appear at the bottom of the page in red.
- Repeat this process for each primer you are using.

Melting Temperature (Tm) Calculation

1 — **Primer (6-50 bases):**
GTATGTGTGTATATATATGT

Compute Tm — 4

LENGTH	20
C+G%	25
Molecular weight:	6272.715

Basic Tm
Degenerated nucleotides are allowed

2 — **Base-Stacking Tm**
Degenerated nucleotides are NOT allowed

Primer concentration: 500 nM

Salt concentration: 50 mM — 3

Mg²⁺ concentration: 1.5 mM

Tm: 50.7 °C — 5

Enthalpy: -145.6
Entropy: -419.42

Source code is freely downloadable at biophp.org





Submission Instructions:

- Download the submission form from: <https://iki-labs.bgu.ac.il/genomic-analysis-2/> .
- Fill out the form (manual or automatic) and attach it to the samples when submitting.
- Alternatively, you can email the completed form to avishagc@post.bgu.ac.il and shirandr@bgu.ac.il, and submit the samples with a note containing your name, phone number and lab.
- Submit the samples to **building 39, room 205 2nd floor**. In the fridge on the 2nd shelf from the bottom there is a tray with PCR tube stands for your use.
- Outside of working hours, you can submit the samples to the mailbox at the entrance to the unit. When submitting via mailbox, samples must be placed in a closed Ziplock bag with your form\note. Make sure your tubes are sealed!
- Samples are collected from the mailbox at 10:00 (AM) every day from Sunday to Thursday.

Thank you for your cooperation,

Avishag Cahana

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